Solvent Effects on the UV−Vis Absorption Properties and Tautomerism of N‑Confused Tetraphenylporphyrin

Elvin A. Alemán, *^{,†} Jojo Joseph,[§] and David A. Modarelli^{*,§}

§ Department of Che[mis](#page-6-0)try and The Center for Laser and Optical Spect[ros](#page-6-0)copy, Knight Chemical Laboratory, The University of Akron, Akron, Ohio 44325-3601, United States

† Department of Chemistry, California State University Stanislaus, One University Circle, Turlock, California 95382, United States

ABSTRACT: The two tautomeric forms of N-confused tetraphenylporphyrin (NCTPP) show distinctly different absorption spectra. The existence of each tautomer in solution has been shown to be strongly solvent-dependent. In the present work, we have studied the tautomerization using absorption spectroscopy in 15 different solvents. While changes in the two tautomers are not large in the Soret band region, the distinct spectral changes between the two tautomers in the Q-band region provide a convenient way to measure the concentration of each tautomer. The resulting data shows a strong correlation between the tautomer and the H-bond accepting ability of the solvent. The anomaly in this data is for the alcoholic solvents ethanol and methanol, for which we observe evidence for H-bonding, presumably between the exterior N2 nitrogen of the NCTPP and the O−H proton of the solvent.

■ INTRODUCTION

Photosynthesis is initiated by the absorption of light by the bacteriochlorophylls present in the light harvesting (LH) complexes of green plants and purple photosynthetic bacteria. The resulting excitation energy is subsequently transferred to the photosynthetic reaction center (RC), resulting in charge separation across the photosynthetic membrane that is localized between the cytoplasmic and periplasmic membranes.¹ The antenna complex 2 in purple photosynthetic bacteria consists of two types of LH complexes, LH1 and LH2, that surrou[n](#page-6-0)d the RC and togeth[er](#page-6-0) form the so-called photosynthetic unit (PSU).3,4 The major light-absorbing species in all of these assemblies is bacteriochlorophyll a (Bchl a) and its derivatives. The n[ear](#page-6-0)-unity conversion of solar energy into chemical potential in the LHCs and RC has inspired significant research toward the design and study of molecular systems that effectively mimic the energy- and electron-transfer processes in photosynthesis, with the ultimate goal of efficiently converting solar energy into electricity.

A detailed understanding of the photophysical properties of bacteriochlorophylls and more generally the parent porphyrins has been important for elucidating mechanisms such as energy and electron transfer in biological complexes, as well as artificial photosynthetic analogues, molecular electronics, and photonic devices.^{5,6} Porphyrins contain an 18-electron aromatic ring with four nitrogen atoms in the core of the macrocycle. This conjugated π -system is the electronic "heart" of the macrocycle and is responsible for the intensity, color, and optical properties of porphyrins. Changes to the tetrapyrrolic macrocycle upon reduction of the pyrrole rings, $\frac{7}{8}$ fusing different rings across the pyrrole rings of the macrocycle,^{9−11} or exchanging, adding, or removing atoms or molecula[r g](#page-6-0)roups in the macrocycle, often result in substantial c[hang](#page-6-0)es in the photophysical properties. The photophysical properties of free [b](#page-6-0)a[se](#page-6-0) tetraphenylporphyrin $(H_2TPP,$ Scheme 1) are well understood, and the molecule, as a result, is often used as standard to study the photophysical propertie[s of oth](#page-1-0)er porphyrins. The absorption spectra of free base tetraarylporphyrins typically show four relatively weak bands in the 480−800 nm region that are collectively called the Q-bands, and correspond to the forbidden $S_0 \rightarrow S_1$ transition. A more intense band is observed at higher energy (390−450 nm) that results from the strongly allowed $S_0 \rightarrow S_2$ transition (the B-band or Soret band).

N-Confused tetraphenylporphyrins 1e and 1i (Scheme 1) are isomers of tetraphenylporphyrin that have one pyrrolic nitrogen exchanged with one of the pyrrolic β -carbon[s so that](#page-1-0) one nitrogen faces outside the macrocycle.^{17,18} Three possible

Received: September 25, 2015 Published: October 12, 2015

Scheme 1

tautomers of NCTPP have been postulated to coexist in equilibrium,¹⁹ although only two of the tautomers have been observed experimentally.17,18,20 The most stable tautomeric form (1i) h[as](#page-6-0) three hydrogens inside the macrocycle, with the outer nitrogen unproton[ated, w](#page-6-0)hile the second tautomer (1e) has two hydrogens inside the ring and one on the external nitrogen.

We have previously investigated the photophysical properties of unsubstituted and substituted NCTPPs 1i and 1e in solution using a combination of computations, 21 steady-state fluorescence, time-resolved fluorescence, and ultrafast absorption spectroscopies. $22,23$ These properties [ar](#page-7-0)e of great interest because of the potential for incorporation of NCTPPs into applications fo[r whi](#page-7-0)ch porphyrins are normally considered. The absorption spectra of the tautomers are quite different from one another. Tautomer 1i (in CH_2Cl_2) exhibits a Soret band at 437 nm and has a Q-band structure similar to that observed for freebase tetraphenylchlorin (H_2TPC) ,⁷ with a maximum Q-band absorption at 724 nm. Tautomer 1e (in DMAc), on the other hand, has a Soret band absorpti[on](#page-6-0) at 442 nm and Q-band profile which gains in intensity with increasing wavelength to a maximum at 699 nm.

 Furuta^{20} and Latos-Grazynski 17 presented the first direct evidence for the tautomerization of 1i and 1e, and Furuta suggeste[d t](#page-6-0)he equilibrium was p[os](#page-6-0)sibly dependent on solvent polarity.²⁰ Absorption experiments showed a dramatic change in both the color and the spectra of solutions of NCTPP in CH_2Cl_2 [\(r](#page-6-0)ed) and DMF (green). Using ${}^{1}H$ NMR and X-ray diffraction analysis, Furuta and co-workers determined that the tautomeric form 1e predominates in DMF, while 1i is observed in CH_2Cl_2 .²⁰ Furuta also observed the presence of both tautomers in pyridine, resulting in a proposed mechanism for the transiti[on](#page-6-0) from 1i to 1e wherein the confused pyrrole ring buckles toward the opposing core nitrogen, leading to proton exchange and subsequent tautomerization.^{20,24,25}

A variety of subsequent experiments support the solvent dependent tautomerization, and several g[ro](#page-6-0)[ups h](#page-7-0)ave used this information to study the various properties of 1e vs 1i, as well as the tautomeric forms of other NCTPP derivatives.^{22,23,26−28} Recently, the construction of self-assembled supramolecular systems was investigated as a function of NCTPP [tautomer,](#page-7-0) and a proposed mechanism of tautomeric structures in a combination of four protic and aprotic solvents using SEM and AFM analysis was described.²⁹ Computational work by Marchand and Jacquemin, 30 and separately by us, $21,31$ using the polarization continuum mo[del](#page-7-0) in Gaussian09 and density functional theory (DFT), [con](#page-7-0)firmed energetic diffe[rence](#page-7-0)s as a function of solvent polarity, but also showed a switch in tautomer preference did not occur under these conditions. We

previously examined the solvent dependent nature of the tautomerization by UV−vis spectroscopy, confirming Furuta's observations that both NCTPP tautomers can exist simultaneously in certain solvents, presumably in equilibrium with one another. 32 In the present work, we extend this original analysis and present a more detailed photophysical investigation of the 1i/1e t[aut](#page-7-0)omerism using a wide range of solvents in order to correlate the dependence of the NCP tautomerism with one or more of the following solvent properties: polarity/dipolarity, Hbond donor propensity, H-bond acceptor propensity, dipole moment, and dielectric constant.^{33,34}

■ RESULTS AND DISCUSSI[ON](#page-7-0)

Gu et al.³⁴ have previously applied a mathematical model to scale the parameters for solvent polarity/dipolarity (π) , the Hbond do[nor](#page-7-0) $(\Sigma \alpha)$, and the H-bond acceptor $(\Sigma \beta)$ properties to a common scale of zero to one. These values are shown in Table 1, together with the literature values for dipole moment and dielectric constant for each solvent.³⁴ Because previous descriptions of this tautomerism have invoked solvent polarity as the main factor governing the directio[n o](#page-7-0)f the equilibrium, we have organized this table so the other parameters are tabulated relative to the solvent polarity function, π .

Table 1. Solvent Parameters for the Solvents Used To Study the NCTPP Tautomerism^{33,34}

solvent	π^a	Σa^b	$\Sigma \beta^c$	dipole moment (D)	dielectric constant
DMSO	1.00	0.00	0.88	3.96	46.83
Benzonitrile	0.90	0.00	0.33	4.18	25.59
DMAc	0.88	0.00	0.78	3.70	37.78
Pyridine	0.87	0.00	0.52	2.22	12.98
Dichloromethane	0.82	0.10	0.05	1.60	8.93
Dichloroethane	0.81	0.10	0.11	1.80	10.13
Acetonitrile	0.75	0.07	0.32	3.92	35.69
Acetone	0.71	0.04	0.49	2.88	20.49
Butyronitrile	0.71	0.00	0.36	4.07	24.29
Methanol	0.60	0.43	0.47	1.70	32.61
Chloroform	0.58	0.15	0.02	1.04	4.71
THF	0.58	0.00	0.48	1.75	7.43
Ethanol	0.54	0.37	0.48	1.69	24.85
Toluene	0.54	0.00	0.14	0.38	2.37
Cyclohexane	0.00	0.00	0.00	0.00	2.02

a The solute dipolarity−polarizability term reported by Abraham represents the dipole−dipole or dipole-induced−dipole plus some polarizability interaction in the solute-condensed phase interaction.³³
^bHydrogen bond donor propensities of the solvent. ^cHydrogen bond acceptor propensities of the solvent.

Figure 1. Steady-state absorption spectra of NCTPP tautomers 1i and 1e in the 300−500 nm range in a series of solvents. The spectra of NCTPP in DMSO (black) and in cyclohexane (gray) are shown together with the spectrum in the solvent (red) under investigation.

Dimethyl sulfoxide (DMSO) and cyclohexane represent the two extremes in terms of polarity, hydrogen bond acceptor value, and dielectric constant. DMSO has the highest polarity $(\pi \sim 1.00)$, the highest hydrogen bond acceptor value (Σ $\beta \sim$ 0.88), and the greatest dielectric constant (46.83) of the solvents examined in this work, while cyclohexane has the lowest polarity (0.00), hydrogen bond acceptor value (0.00), and dielectric constant (2.02). To compare the tautomerism of 1i/1e in each solvent used, it was assumed that the steady-state absorption spectrum in DMSO corresponded to 100% of tautomer 1e, while the spectrum in cyclohexane corresponded to 100% of tautomer 1i. The other solvents in Table 1 represent a range intermediate between these two extremes, allowing the analysis of several criteria that might inf[orm what](#page-1-0) solvent-related factor(s) specifically influence the interconversion between the two tautomers. The steady-state absorption spectra of NCTPP in each of the solvents used, compared to the spectra of DMSO and cyclohexane, are shown in Figure 1 (Soret or B-band) and Figure 2 (the significantly more diagnostic Q-band region), while the absorption data from these experiments in bot[h the 300](#page-3-0)−500 nm (B-band) and 500−850 nm (Q-band) ranges are listed in Tables 2 and 3, respectively. The log of the apparent extinction coefficients (log

 ϵ) for each wavelength, which correspond to either one tautomer or a contribution from both tautomers absorbing at that wavelength, are also shown in Tables 2 and 3.

The B-band region (300−500 nm) in DMSO displays a maximum absorption wavelength (λ_{max}) at 444 [nm](#page-4-0) (log $\varepsilon =$ 5.20), while in cyclohexane a value of 436 nm (log ε = 5.32) is observed. In general, the maximum absorption wavelengths in this region for solvents with very high polarities (>0.80) and high hydrogen bond acceptor values (>0.50) were observed in the range of 442−444 nm (DMSO, benzonitrile, DMAc, and pyridine). In solvents such as dichloromethane (CH_2Cl_2) and dichloroethane $(EtCl₂)$ that have high polarity constants (0.80) but low H-bond acceptor values (<0.11), λ_{max} was blue-shifted to 437 and 439 nm, respectively. In acetonitrile, acetone, and butyronitrile, which have high polarity constants (0.71−0.75) and medium of H-bond acceptor values (0.20−0.49), the NCTPP B-band absorption wavelength was very similar to that observed in cyclohexane. In toluene and chloroform, which have low polarities (0.54 and 0.58) and low H-bond acidities (0.00 and 0.15), absorption maxima at wavelengths in between DMSO and cyclohexane were observed.

Methanol and ethanol differ from the previously mentioned solvents because they are the only solvents examined that have

Figure 2. Steady-state absorption spectra of NCTPP tautomers 1i and 1e in the 500−850 nm range in a series of solvents. The spectra of NCTPP in DMSO (black) and in cyclohexane (gray) are shown together with the spectrum in the solvent (red) under investigation.

Table 2. Steady-State Absorption Data for NCTPP in the 300−500 nm Range in a Series of Solvents

	Absorption/nm ($\log \epsilon$)					
solvent	N-band	shoulder ^{<i>a</i>}	B-band	shoulder ^b		
DMSO	359(4.58)	425 (4.89)	444(5.20)	$\overline{}$		
Benzonitrile	393(4.51)	427(5.00)	443(5.33)			
DMAc	357(4.54)	423(4.81)	442(5.08)	476 (4.31)		
Pyridine	362(4.60)	426 (4.96)	444(5.26)			
Dichloromethane	393 (4.36)	422(4.86)	438 (5.21)			
Dichloroethane	383 (4.37)	424 (4.90)	439(5.20)			
Acetronitrile	359 (4.52)		435(5.28)	469(4.45)		
Acetone	363(4.23)		437(5.01)	471 (4.27)		
Butyronitrile	360(4.46)	421 (4.93)	436 (5.19)			
Methanol	388 (4.46)	416 (4.86)	437(5.24)			
Chloroform	393(4.30)		440 (5.14)			
THF	363(4.48)	423(5.02)	438 (5.27)			
Ethanol	383(4.33)	419 (4.86)	436(5.20)			
Toluene	390 (4.64)	$\overline{}$	441 (5.44)			
Cyclohexane	393 (4.60)	423(5.06)	436(5.32)			

 a The shoulder to the blue of the B-band. b The shoulder to the red of the B-band.

moderately high H-bond donor values (0.43 and 0.37, respectively). As a result, if solvent acidity plays any kind of role in the tautomerization, it should show the most dramatic influence in these solvents. In addition, these two solvents have polarity values similar to chloroform and H-bond acceptor values comparable to acetone. Despite the relatively high Hbond donor and polarity values of these solvents, the observed B-band of NCTPP is the same as that observed in cyclohexane (1i), indicating the H-bond donor ability $(\Sigma \alpha)$ of the solvent does not seem to influence the equilibrium. In THF, which is similar in polarizability and H-bond accepting ability to methanol and ethanol, the B-band absorption wavelength occurs at ∼438 nm, similar to the absorption band energy observed in cyclohexane. These results indicate that tautomer 1i is favored in solvents with H-bond acceptor values lower than 0.50. When NCTPP is dissolved in a solvent having a higher H-bond acceptor value, such as DMSO or DMAc, the band absorption maxima is red-shifted to that of 1e. The outlying data point in this analysis is benzonitrile, which has a high polarity value (0.90) and low H-bond acceptor value (0.33), and for which a value of $\lambda_{\rm max}$ of 443 nm was observed. However, the shape of the B-band in benzonitrile is similar to that in cyclohexane, which suggests the red-shift in the B-band in this solvent is related to other factors, such as refractive index, 35 that are not considered in this analysis. In general, the ratio of the two tautomeric forms does not appear to appre[cia](#page-7-0)bly depend on solvent polarity.²⁰

The close overlap between the Soret absorption maxima of the two tautomers in different solven[ts](#page-6-0) makes it difficult in solvents intermediate between cyclohexane and DMSO to definitively identify the ratio of tautomers. The Q-band region (500−750 nm), in which each tautomer has a distinct absorption pattern, was therefore used to determine quantitatively the amount of each NCTPP tautomer in each solvent. In cyclohexane, two high-energy Q-bands for 1i are observed as very intense absorptions at 540 and 581 nm, respectively, whereas a third, lower-energy Q-band at 665 nm is quite weak. An intense low energy Q-band for 1i is observed at 724 nm. In DMSO, the high-energy, low intensity Q-bands for 1e at 556 and 596 nm overlap the 540 and 581 nm Q-bands of 1i, and cannot easily be used to distinguish between the tautomers. However, the low-energy Q-bands for 1e at 645 and 699 nm are both quite intense, and conveniently occur in a region of the spectrum where 1i does not have a strong

absorption. As a result, this region of the spectrum provides a window where quantitative comparisons between 1i and 1e are possible. The red side of the 699 nm band of 1e overlaps somewhat the blue side of the 724 nm band of 1i, and analysis of these bands to quantify the amount of each tautomer in those solvents where both tautomers coexist was accomplished using eqs 1 and 2. Because the differences in the H-bond donor values are quite small, except in the case of methanol and ethanol, we have largely limited the following discussion to just examine the Q-band ratios of 1i and 1e as a function of the Hbond acceptor values and polarity values.

The Q-band region in the absorption spectra in DMSO and DMAc are very similar to one another, indicating the same tautomer (1e) is present in both solvents. In solvents such as pyridine, acetonitrile, acetone, and butyronitrile, which have high polarity values but lower H-bond acceptor values, the absorption spectra showed evidence of the presence of both tautomers. To approximate the quantity of each tautomer, it was assumed that the changes in the extinction coefficients of the Q-bands for 1e and 1i as a function of solvent were minor.^{36,37} By measuring the absorption at ∼581 nm (where 1i absorbs significantly more strongly than 1e) and ∼645 nm (wher[e](#page-7-0) [1e](#page-7-0) absorbs more intensely than 1i), the relative percent of each tautomer in solution was estimated as follows (eqs 1 and 2):

$$
\frac{n_{1i}}{V_{T}} = \frac{\begin{vmatrix} A_{645} & A_{581} \\ \varepsilon_{645}^{1e} & \varepsilon_{581}^{1} \\ \varepsilon_{645}^{1e} & \varepsilon_{581}^{1} \\ \varepsilon_{645}^{1e} & \varepsilon_{581}^{1} \\ \varepsilon_{645}^{1e} & \varepsilon_{581}^{1} \end{vmatrix}} \qquad \frac{n_{1e}}{V_{T}} = \frac{\begin{vmatrix} A_{645} & A_{581} \\ \varepsilon_{645}^{1i} & \varepsilon_{581}^{1} \\ \varepsilon_{645}^{1e} & \varepsilon_{645}^{1e} \\ \varepsilon_{581}^{1e} & \varepsilon_{645}^{1e} \\ \varepsilon_{581}^{1e} & \varepsilon_{645}^{1e} \\ \varepsilon_{581}^{1e} & \varepsilon_{645}^{1e} \end{vmatrix}}{\begin{vmatrix} \varepsilon_{1}^{1e} & \varepsilon_{1}^{1e} \\ \varepsilon_{645}^{1e} & \varepsilon_{645}^{1e} \\ \varepsilon_{781}^{1e} & \varepsilon_{645}^{1e} \\ \varepsilon_{811}^{1e} & \varepsilon_{645}^{1e} \end{vmatrix}} (1)
$$

$$
\% \mathbf{1i} = \frac{n_{1i}}{n_{1i} + n_{1e}} \times 100 \qquad \% \mathbf{1e} = \frac{n_{1e}}{n_{1i} + n_{1e}} \times 100
$$
\n(2)

where A_x corresponds to the absorption at a given wavelength (x) in each solvent, ϵ_x is the extinction coefficient determined at that same wavelength for 1e in DMSO and 1i in cyclohexane, and n represents the moles of each tautomer in solvents where both NCTPP tautomers are present simultaneously. With this analysis, the amount of the externally protonated tautomer 1e in pyridine (∼69%), acetone (∼64%) and butyronitrile (∼51%), and acetonitrile (∼46%) shows a better correlation

Figure 3. Correlation of 1e formation with the solvent properties (a) diaelectric constant, (b) H-bond acceptor propensities, and (c) polarity/ dipolarity.

Figure 4. Simulated steady-state absorption spectra of NCTPP tautomers 1i and 1e in the 500−850 nm range in a series of solvents. The spectra of NCTPP in DMSO (black) and in cyclohexane (gray) are shown to compare the simulated spectrum in the solvent (red) under investigation. The simulated spectra (blue) were plotted as a function of the percent contribution of each tautomer.

with the trend in the H-bond acceptor value (pyridine (0.52)) acetone (0.49) > butyronitrile (0.36) > acetonitrile (0.32)) than with the trend in polarity (pyridine (0.87)) acetonitrile (0.75) > acetone (0.71) ~ butyronitrile (0.71)). In pyridine and acetone, which have similar H-bond acceptor values (0.52 and 0.49) but relatively larger differences in polarity (0.87 and 0.71), the ratio of 1e to 1i is nearly equivalent. However, a different ratio of 1e to 1i is observed in acetone (64% 1e) and butyronitrile (51% 1e), both of which have the same polarity values (0.71) but different H-bond acceptor values (0.49 and 0.36). This data is summarized in Figure 3.

The Q-band regions in the absorption spectra of NCTPP in CH_2Cl_2 and EtCl₂, both of which have very high polarities (0.82 and 0.81) and very low H-bond acceptor values (0.05 and 0.11), are near-perfect matches with the absorption spectrum of NCTPP in cyclohexane. Similarly, the Q-band region in CHCl₃ and toluene, both of which have a very low H-bond acceptor value (0.02 and 0.14) but moderate polarity (0.58 and 0.54), indicates the major tautomer in either solvent is 1i. In benzonitrile, which has a very high polarity value (0.90) and a low-to-moderate H-bond acceptor value of 0.33, analysis of the absorption spectrum indicates the concentration of 1i is quite high (81%) but that the equilibrium does not completely favor its formation, as is observed for the halogenated solvents or toluene. THF has a significantly lower polarity value of 0.58 but a higher H-bond acceptor value (0.48) than benzonitrile. The change in the H-bond acceptor value is to push the equilibrium more toward 1e, thereby decreasing the concentration of 1i in this solvent to ∼63%. From these results, the tautomerism of NCTPP in solution apparently shows a strong dependence on

the H-bond acceptor of the solvent and very little dependence on the polarity (Figure 3). The absorption spectra were simulated 38 in the Q-band region with good correlation using the relative amount of each tautomer, shown in Figure 4 as the percent c[on](#page-7-0)tribution relative to the standard spectra of DMSO and cyclohexane.

The effect of solvent on the related tautomerization within a series of keto−enol pairs has been previously investigated by Mills and Beak 39 using a similarly wide range of solvents. In their work, the keto−enol equilibrium was also found to primarily be de[ter](#page-7-0)mined by the hydrogen-bonding basicity (Hbond accepting ability) of the solvent. The enol forms were observed to be preferentially stabilized by H-bonding-acceptor solvents, and when the correlations were calculated, the whole weight of the correlations were carried by the β term, which corresponds to the H-bonding accepting property of each solvent. The same correlations were obtained when the dielectric constant term was deleted in the analysis. Therefore, the major interaction of the isomers with the surrounding is the ability of the enolic forms to act as a hydrogen-bond donor to a basic solvent. We believe the same effect is operational in the NCTPP tautomerization.

Furuta et al. have previously proposed 24 a mechanism for the tautomerization wherein the outer nitrogen (N2) on the A-ring, which the crystal structure shows 20 is [alr](#page-7-0)eady rotated out-ofplane toward the two interior N−H groups in the B- and Drings, initiates the tautomerization [in](#page-6-0) an intramolecular fashion by removing one of the inner N−H protons. The formation of tautomer 1e in the Furuta mechanism results from the preferred stability of this tautomer in a polar solvent. The

Furuta mechanism is also consistent with the H-bond accepting solvent results described here, where 1e is more stable in Hbond accepting solvents, biasing the equilibrium in this direction. A second alternate mechanism can also be proposed for the tautomerization that involves two or more solvent molecules. In this mechanism, one H-bond accepting solvent molecule is hydrogen bonded to one of the inner N−H bonds in the B- or D-rings, and, together with a second solvent that adds a proton to the external nitrogen, catalyzes the tautomerization. This reaction would occur in a concerted or near-concerted process similar to many deprotonation− protonation reactions present in biological systems. Furuta et al. have examined the equilibrium in pyridine- d_5 and observed ΔG^{\ddagger} , ΔH^{\ddagger} , and ΔS^{\ddagger} values of 14.94 kcal mol⁻¹, 1.93 kcal mol⁻¹, and −47.6 eu, respectively.²⁰ The low activation enthalpy is consistent with a concerted reaction, and the rather negative entropy of activation with the highly ordered transition state predicted by an intramolecular mechanism. For reference, Abraham and co-workers reported values of 13.5 kcal mol[−]¹ , 9.2 kcal mol⁻¹, and −10 eu for the NH shift in H₂TPP. Importantly, the transition state for the H_2TPP isomerization is not likely to involve the out-of-plane distortion/rotation required by Furuta's NCTPP mechanism.⁴⁰

Interestingly, the absorption spectra acquired in methanol and ethanol are also consistent with the [pre](#page-7-0)sence of 1i (i.e., in cyclohexane). These solvents, which have the highest H-bond donor values (0.43 and 0.37, respectively) and moderate Hbond acceptor values (0.47 and 0.48), shift the equilibrium toward 1i, consistent with a mechanism involving H-bonding of the alcohol to the lone pair of electrons on the external nitrogen. Several groups 17,18,26 have determined the absorption spectra of NCTPP under acidic conditions, and have shown both tautomers undergo se[que](#page-7-0)ntial mono- and diprotonations, a process that is accompanied by new absorption bands at ∼790 nm for the monocation and ∼840 nm for the dication in CHCl₃, and ~840 nm in DMF.^{18,26,27} The absorption spectra in both solvents, and in particular methanol, show a low energy tail to the red of the $Q_v(0,0)$ band [that](#page-7-0) extends toward 800 nm. We suggest this tail results from H-bonding from the solvent to the exterior nitrogen (N2). From the absorption spectra, it is clear the proton is not fully shifted to the nitrogen, a result that is not surprising given the expected difference in pK_a values between the alcohols and protonated 1i. Nonetheless, the high H-bond acidity in methanol and ethanol may result in Hbonding interactions between the solvent molecules and the external nitrogen of 1i that stabilizes this tautomer, and in any case influences the tautomerization more than the H-bond accepting capacity of the solvent.

■ CONCLUSIONS

The tautomerization of NCTPP was studied in a series of 15 solvents, and the dependence of the equilibrium was examined as a function of several solvent parameters, including polarity/ dipolarity (π) , H-bond donor $(\Sigma \alpha)$, and H-bond acceptor $(\Sigma \beta)$ values; dielectric constant; and dipole moment. The absorption spectra of NCTPP observed in these solvents indicated the tautomerization correlated most strongly with the H-bond acceptor properties of the solvent. In the case of methanol and ethanol, the opposite effect is observed, with evidence of hydrogen-bonding between the exterior N2 nitrogen and the solvent observed.

EXPERIMENTAL SECTION

Absorption spectra were measured on a double-beam spectrophotometer. All solvents used for spectroscopic measurements were either HPLC or spectophotometric grade and were used as received, except chloroform, dichloromethane, and dichloroethane, which were passed through a short column of silica gel prior to use to remove HCl. Every experiment was performed at least three times for reproducibility.

■ AUTHOR INFORMATION

Corresponding Authors

*E-mail: dmodarelli@uakron.edu

*E-mail: ealeman@csustan.edu

Notes

The auth[ors declare no compe](mailto:ealeman@csustan.edu)ting financial interest.

■ ACKNOWLEDGMENTS

E.A.A. gratefully acknowledges the support of California State University Stanislaus (RSCA Grant, G0106-G711-2013-30951) and the U.S. Department of Education for a GAANN fellowship (P200A000203). D.A.M. gratefully acknowledges the support of the National Science Foundation (CHE-1412362, CHE-0526864, CHE-0216371), the Ohio Board of Regents, and The University of Akron. The authors also thank Dr. Shana Garrison for providing samples of NCTPP for use in these experiments, and Mr. Michael Davis of The College of Wooster for his help with the initial absorption experiments.

■ REFERENCES

(1) Fleming, G. R.; Vangrondelle, R. Phys. Today 1994, 47 (2), 48− 55.

(2) Gest, H.; Blankenship, R. E. Photosynth. Res. 2004, 80, 59−70.

(3) Cogdell, R. J.; Fyfe, P. K.; Barrett, S. J.; Prince, S. M.; Freer, A. A.; Isaacs, N. W.; McGlynn, P.; Hunter, C. N. Photosynth. Res. 1996, 48, 55−63.

(4) Hu, X. C.; Schulten, K. Phys. Today 1997, 50, 28−34.

(5) Gust, D.; Moore, T. A.; Moore, A. L. Acc. Chem. Res. 2001, 34, 40−48.

- (6) Wasielewski, M. R. Chem. Rev. 1992, 92, 435−461.
- (7) Gouterman, M. J. Mol. Spectrosc. 1961, 6, 138−163.

(8) Lin, C.-Y.; Spiro, T. G. J. Phys. Chem. B 1997, 101, 472−482.

(9) Kim, J. B.; Leonard, J. J.; Longo, F. R. J. Am. Chem. Soc. 1972, 94, 3986−3992.

(10) Wasbotten, I. H.; Conradie, J.; Ghosh, A. J. Phys. Chem. B 2003, 107, 3613−3623.

(11) Zhang, Y. H.; Ruan, W. J.; Li, Z. Y.; Wu, Y.; Zheng, J. Y. Chem. Phys. 2005, 315, 201−213.

(12) Barkigia, K. M.; Berber, M. D.; Fajer, J.; Medforth, C. J.; Renner, M. W.; Smith, K. M. J. Am. Chem. Soc. 1990, 112, 8851−8857.

- (13) Gentemann, S.; Medforth, C. J.; Forsyth, T. P.; Nurco, D. J.; Smith, K. M.; Fajer, J.; Holten, D. J. Am. Chem. Soc. 1994, 116, 7363− 7368.
- (14) Senge, M. O.; Kalisch, W. W.; Runge, S. Liebigs Ann. Recl. 1997, 1997, 1345−1352.
- (15) Baskin, J. S.; Yu, H. Z.; Zewail, A. H. J. Phys. Chem. A 2002, 106, 9837−9844.
- (16) Yu, H. Z.; Baskin, J. S.; Zewail, A. H. J. Phys. Chem. A 2002, 106, 9845−9854.
- (17) Chmielewski, P. J.; Latos-Grazynski, L.; Rachlewicz, K.; Glowiak, T. Angew. Chem., Int. Ed. Engl. 1994, 33, 779−781.
- (18) Furuta, H.; Asano, T.; Ogawa, T. J. Am. Chem. Soc. 1994, 116, 767−768.
- (19) Szterenberg, L.; Latos-Grazynski, L. Inorg. Chem. 1997, 36, 6287−6291.

(20) Furuta, H.; Ishizuka, T.; Osuka, A.; Dejima, H.; Nakagawa, H.; Ishikawa, Y. J. Am. Chem. Soc. 2001, 123, 6207−6208.

- (21) Vyas, S.; Hadad, C. M.; Modarelli, D. A. J. Phys. Chem. A 2008, 112, 6533−6549.
- (22) Alemán, E. A.; Rajesh, C. S.; Ziegler, C. J.; Modarelli, D. A. J. Phys. Chem. A 2006, 110, 8605−8612.
- (23) Belair, J. P.; Ziegler, C. J.; Rajesh, C. S.; Modarelli, D. A. J. Phys. Chem. A 2002, 106, 6445−6451.
- (24) Furuta, H.; Maeda, H.; Osuka, A. Chem. Commun. 2002, 1795− 1804.

(25) Toganoh, M.; Furuta, H. J. Phys. Chem. A 2009, 113, 13953− 13963.

- (26) Ou, Z.; Chena, X.; Yea, L.; Xuea, S.; Fangb, Y.; Jiangb, X.; Kadish, K. M. J. Porphyrins Phthalocyanines 2015, 19, 251−260.
- (27) Sakashita, R.; Ishida, M.; Furuta, H. J. Phys. Chem. A 2015, 119, 1013−1022.
- (28) Xue, S.; Ou, Z.; Ye, L.; Lu, G.; Fang, Y.; Jiang, X.; Kadish, K. M. Chem. - Eur. J. 2015, 21, 2651−2661.
- (29) Thomas, A. P.; Sreedevi, K. C. G.; Adinarayana, B.; Ramakrishnanb, S.; Srinivasan, A. RSC Adv. 2013, 3, 16967−16972.

(30) Marchand, G.; Roy, H.; Mendive-Tapia, D.; Jacquemin, D. Phys.

Chem. Chem. Phys. 2015, 17, 5290−5297. (31) Vyas, S.; Modarelli, D. A. Unpublished data.

-
- (32) Alemán, E. A. Ph.D. Dissertation, The University of Akron, Akron, OH, 2006.
- (33) Abraham, M. H. Chem. Soc. Rev. 1993, 22, 73−83.

(34) Gu, C. H.; Li, H.; Gandhi, R. B.; Raghavan, K. Int. J. Pharm. 2004, 283, 117−25.

(35) Garcia-Rubio, L. H. Macromolecules 1992, 25, 2608−2613.

(36) Small variations (typically ∼5−10%) in the extinction coefficients of the Q-bands in $H_2 TPP$ as a function of solvent are known. See, for example: Barnett, G. H.; Hudson, M. F.; Smith, K. M. J. Chem. Soc., Perkin Trans. 1 1975, 1401−1403. Datta-Gupta, N.; Malakar, D.; Rice, L.; Rivers, S. J. Heterocycl. Chem. 1987, 24, 629−632. Dudic, M.; Lhoták, P.; Král, V.; Long, K.; Stibor, I. Tetrahedron Lett. 1999, 40, 5949−5952. Kim, J. B.; Leonard, J. J.; Longo, F. R. J. Am. Chem. Soc. 1972, 94, 3986−3992. Thomas, D. W.; Martell, A. E. J. Am. Chem. Soc. 1956, 78, 1338−1343 . It is highly likely that Q-bands in NCTPP also differ somewhat with different solvents. These differences will surely lead to small absolute errors in the 1i/1e ratio calculated for a given solvent.

(37) A similar approach was used to determine the amount of carboxyhemoglobin in blood specimens, where other hemoglobin species were expected to be present: Beutler, E.; West, C. Clin. Chem. 1984, 30, 871−874.

(38) The simulated absorption spectra were calculated using the sum total of the percentage of each tautomer present in solution as a function of the spectra in cyclohexane and DMSO.

(39) Mills, S. G.; Beak, P. J. Org. Chem. 1985, 50, 1216−1224.

(40) Abraham, R. J.; Hawkes, G. E.; Smith, K. M. Tetrahedron Lett. 1974, 15, 1483−1486.